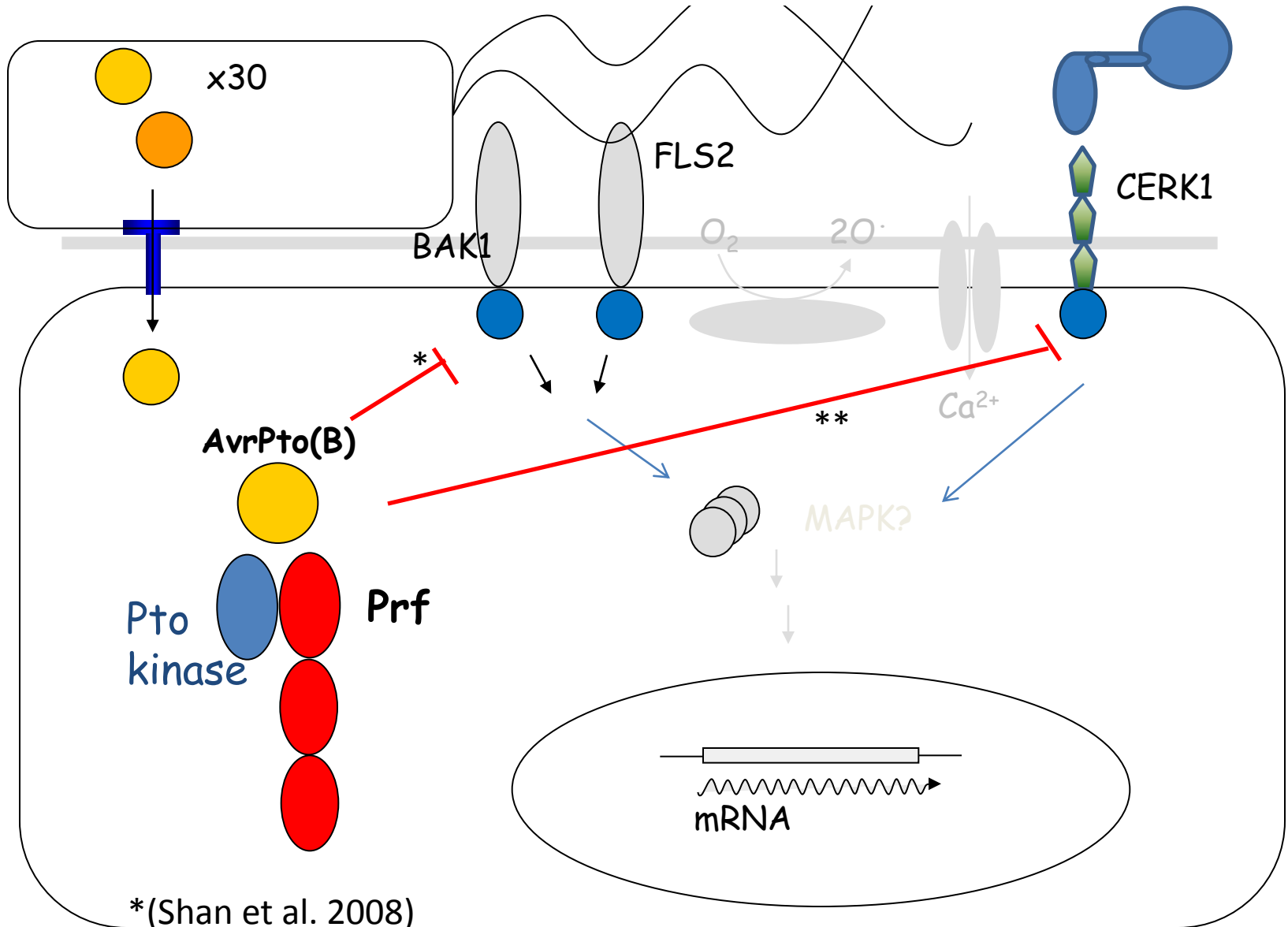


# Moving from identification to quantification of phosphorylation events in plant-pathogen interactions

---



# Effector triggered susceptibility and immunity

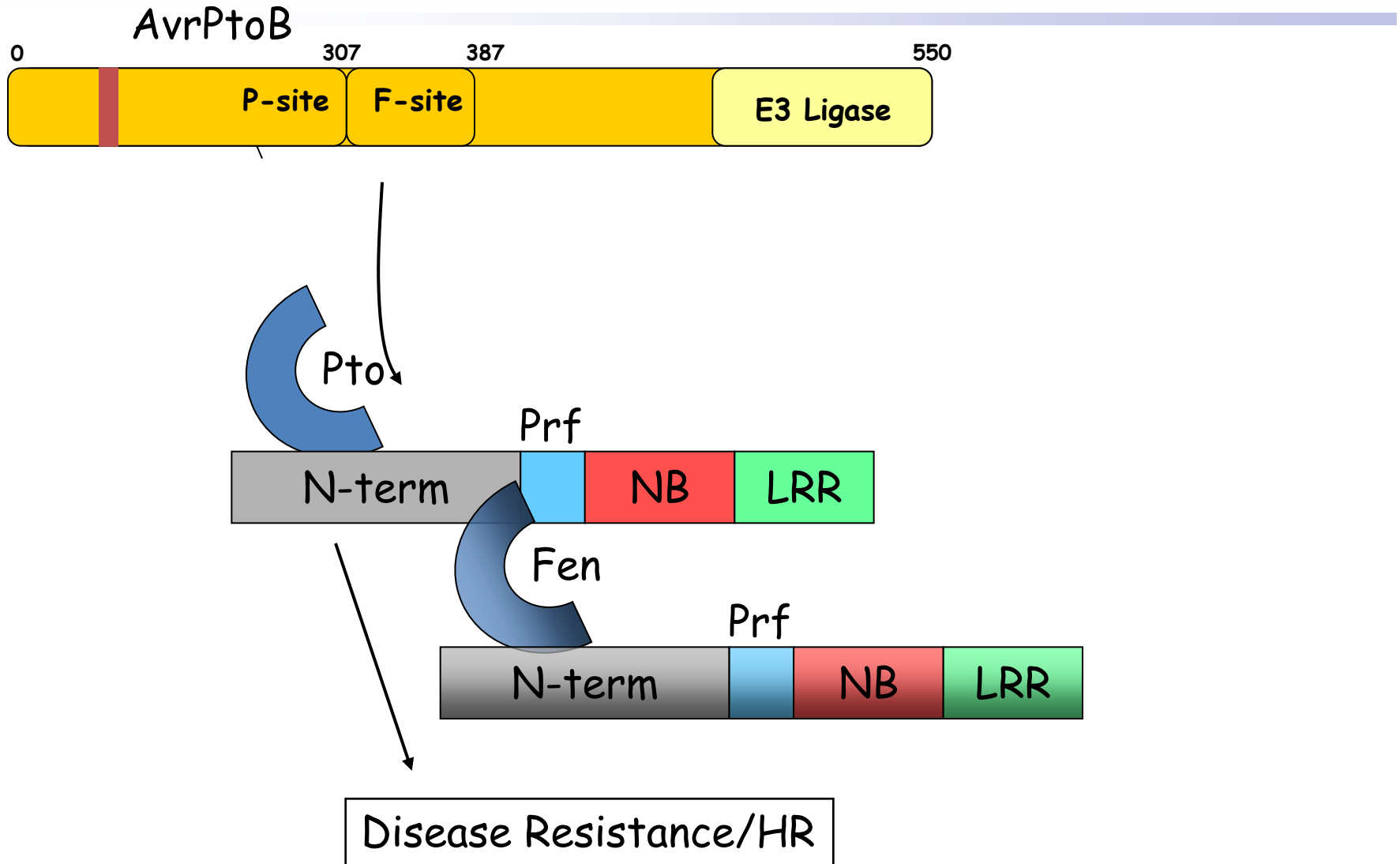


\*(Shan et al. 2008)

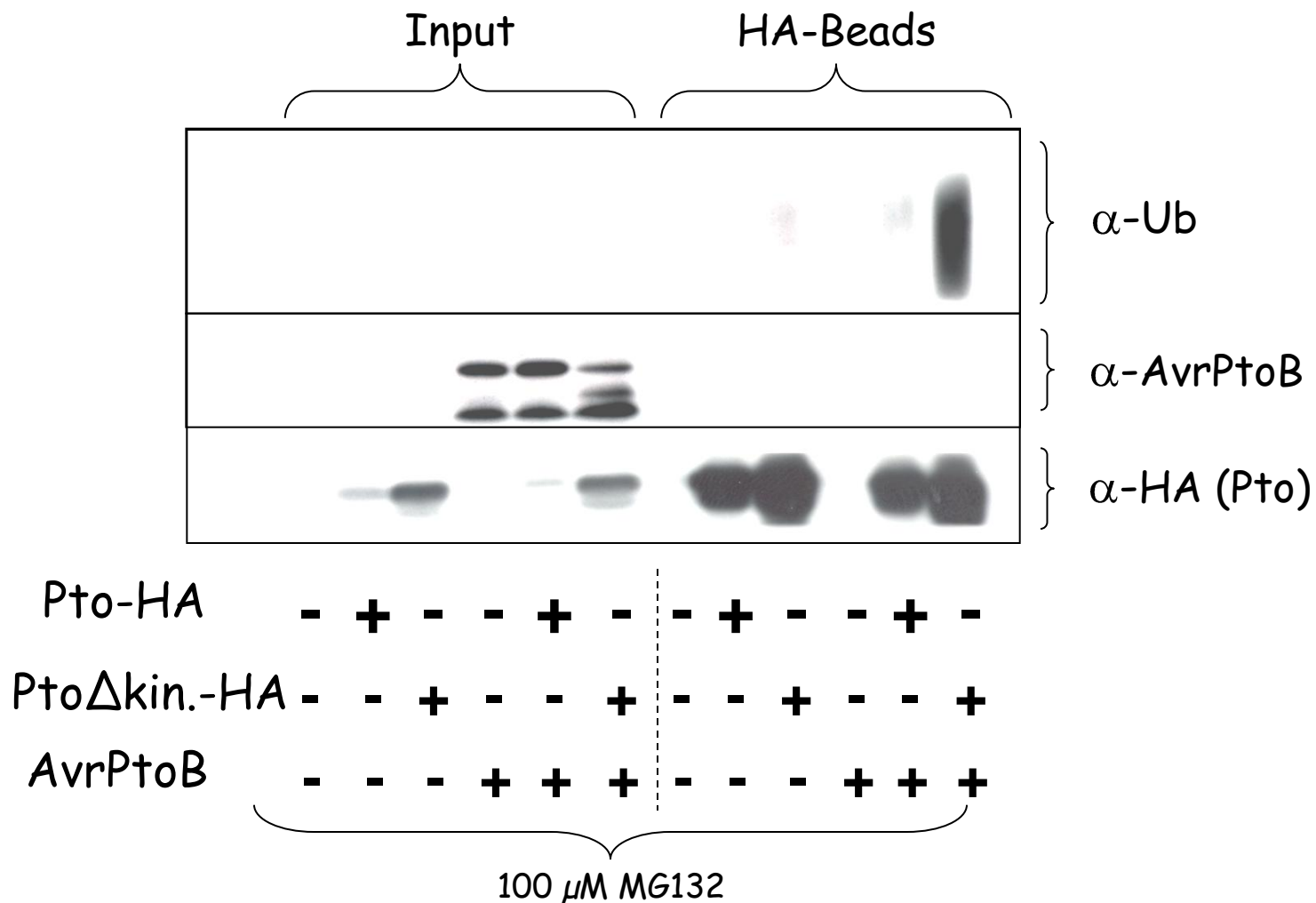
\*\*Gimenez-Ibanez et al Current Biol. 2009

With thanks to John Rathjen for slide

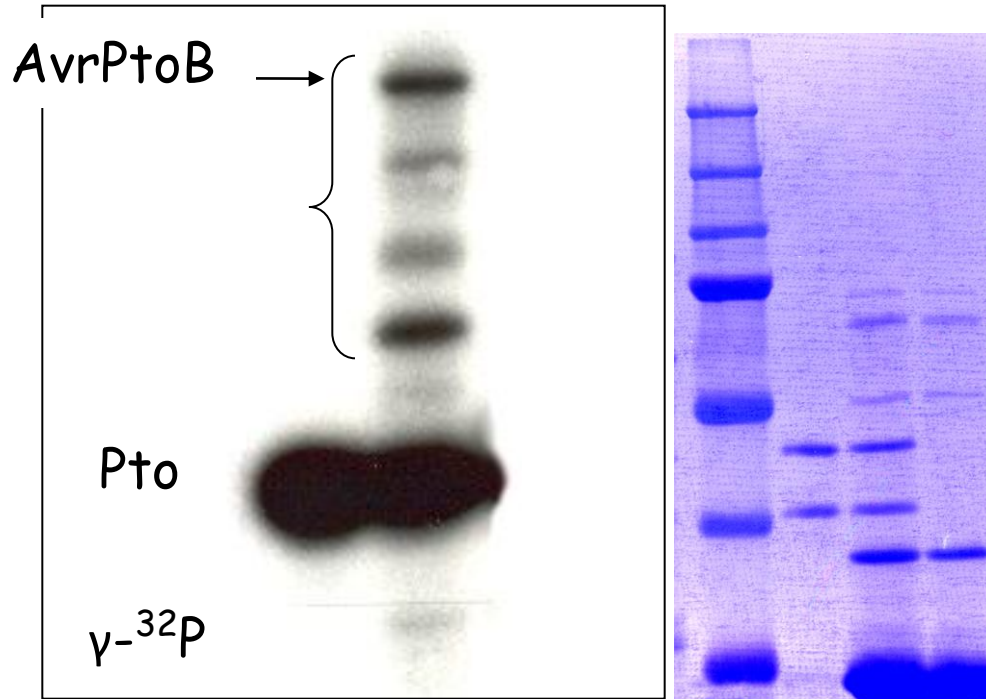
# Signal transduction of AvrPtoB



# AvrPtoB ubiquitinates kinase-dead Pto



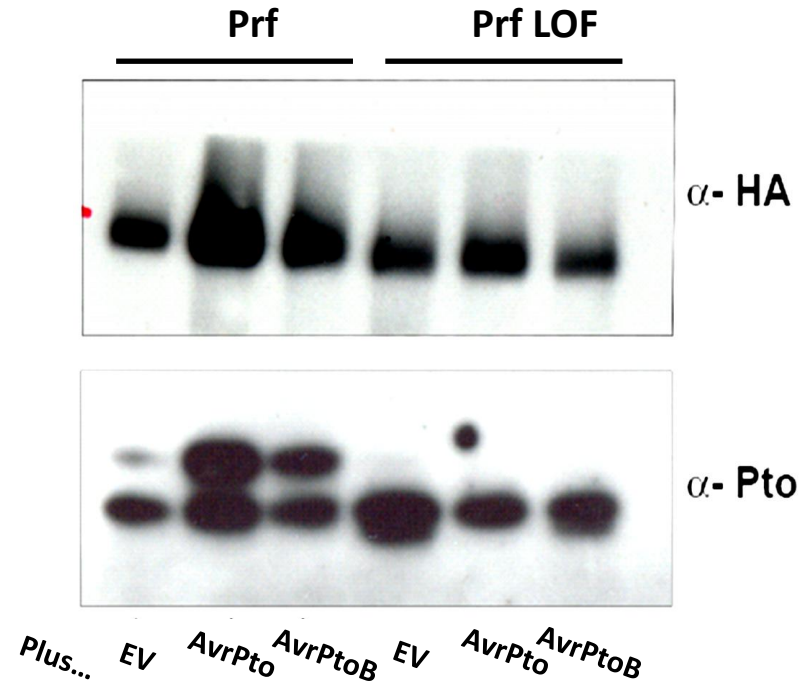
# 1. Pto phosphorylates AvrPtoB



AvrPtoB	-	+	+
Pto	+	+	-

CCB

# 2. Pto is phosphorylated after elicitation



*N. benthamiana*

Ntoukakis et al. Science 2009

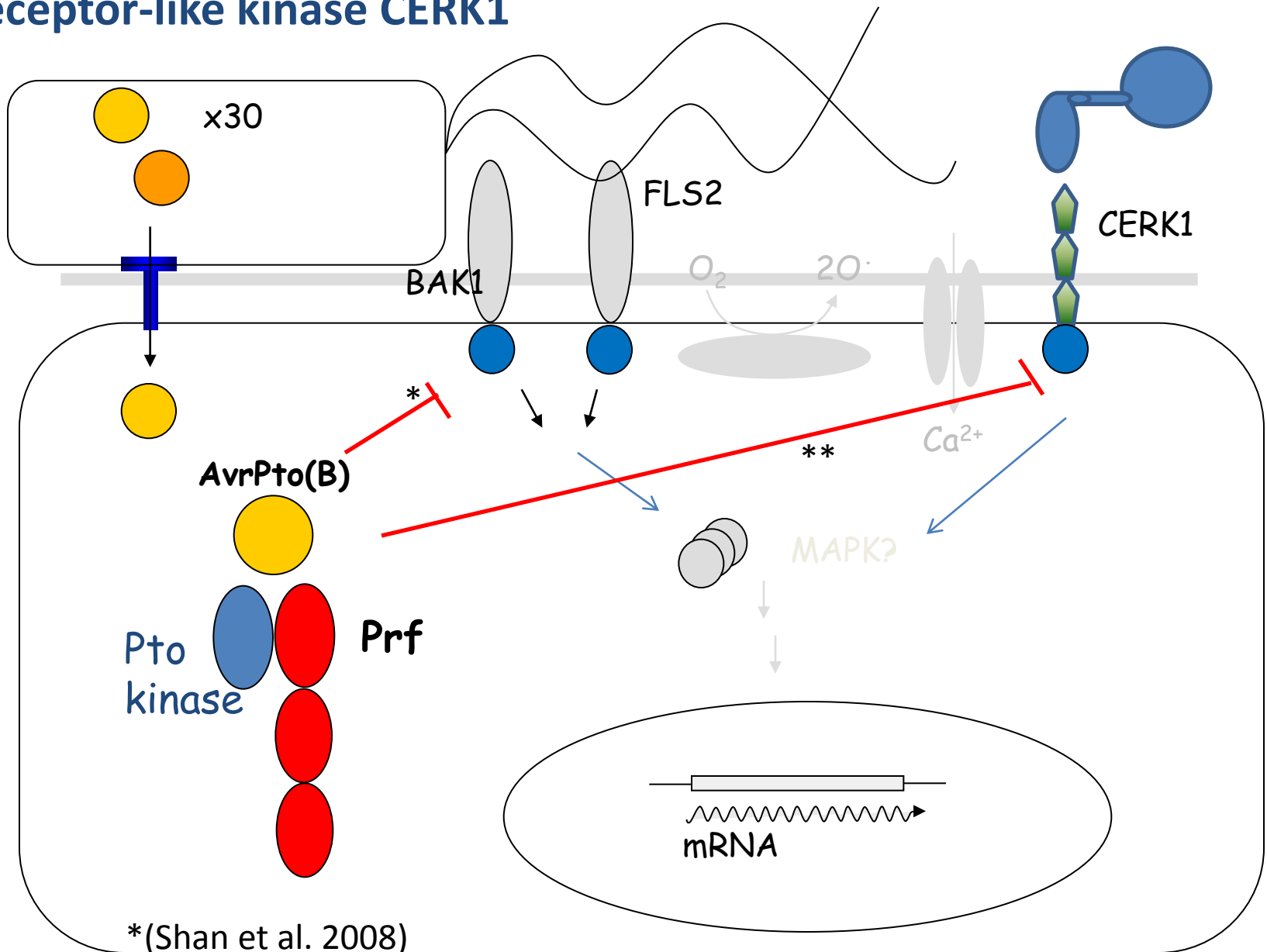
# Modifications on Pto kinase



Proteins: AvrPtoB (purple) Pto pink/yellow.

Space fill: Active site red, 'ubiquitination' site (fen), P-loop residues blue, alternative phosphorylation sites green

# Receptor-like kinase CERK1

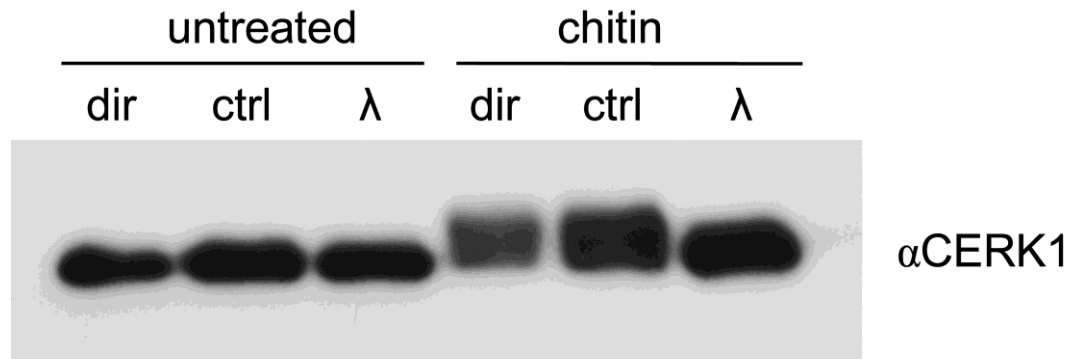
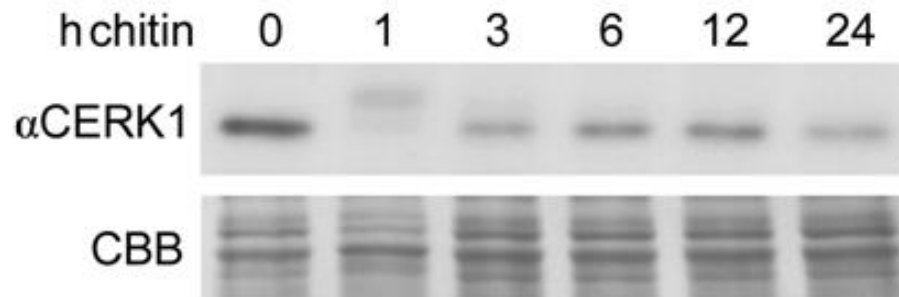
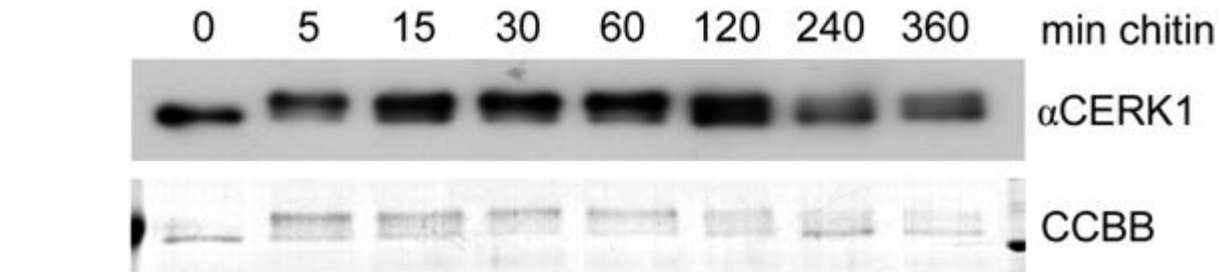


\*(Shan et al. 2008)

\*\*Gimenez-Ibanez et al Current Biol. 2009

With thanks to John Rathjen for slide

# Treatment with chitin induces a transient band-shift



CCBB

Elena Petutschnig



# Identification of constitutive and induced phosphorylation

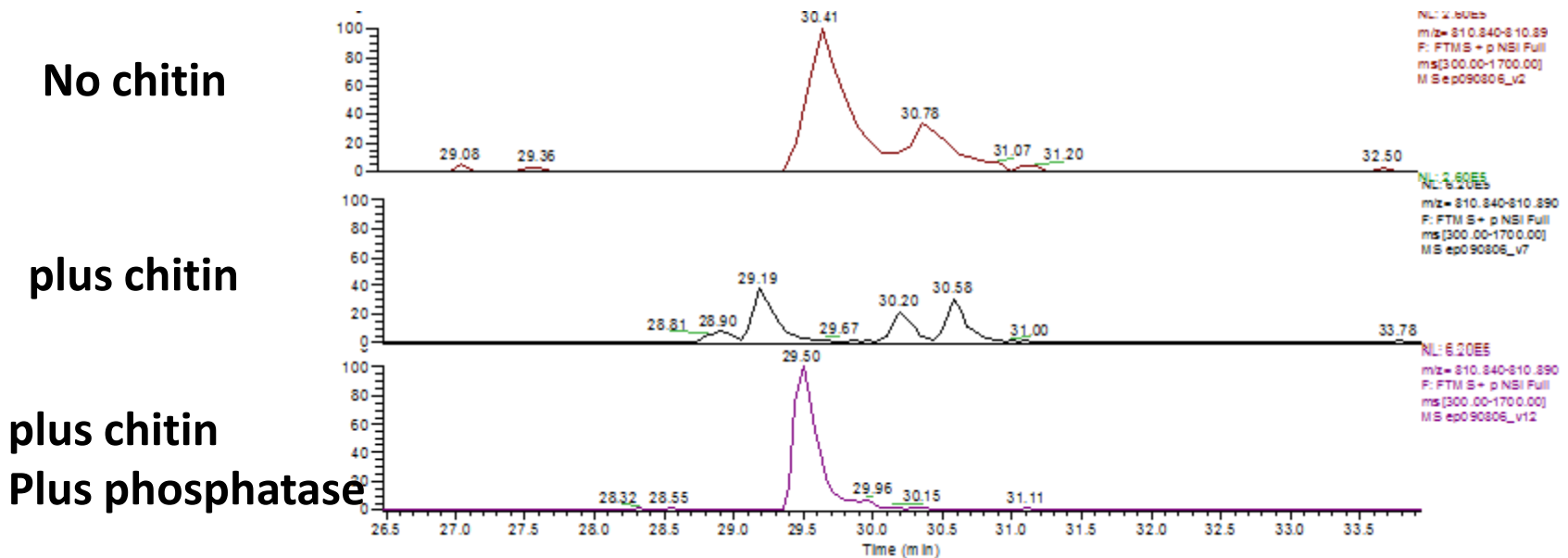
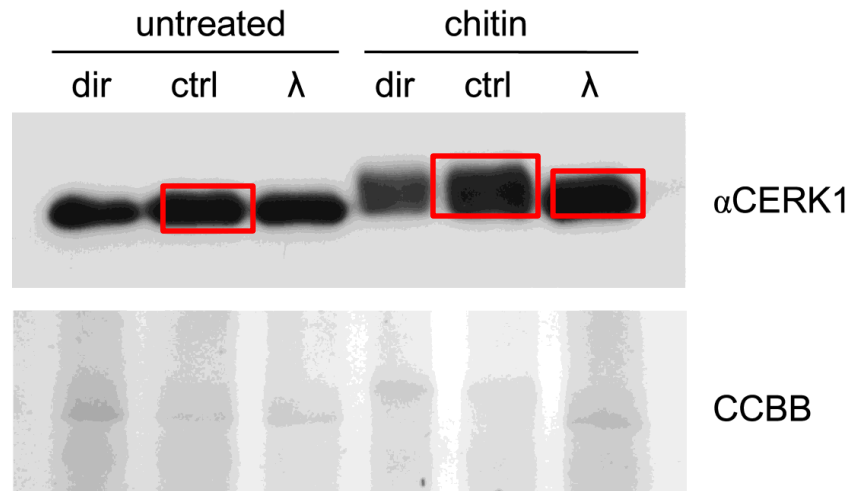
'Quantified' by spectral counts

	Phosphoresidue	control	chitin	dephosphorylated with $\lambda$ -phosphatase	
				control	Chitin
<b>SKGDSFSSSIPLSTK</b>	S266	7	15	1	4
	S268	0	2	0	0
	S270	0	1	0	0
	S274	0	4	0	0
<b>GAVVK[oxM]TEAVGEFR</b>	T519	0	4	0	0

**Two weak points in this analysis:**  
**quantification**  
**position of phosphorylation**

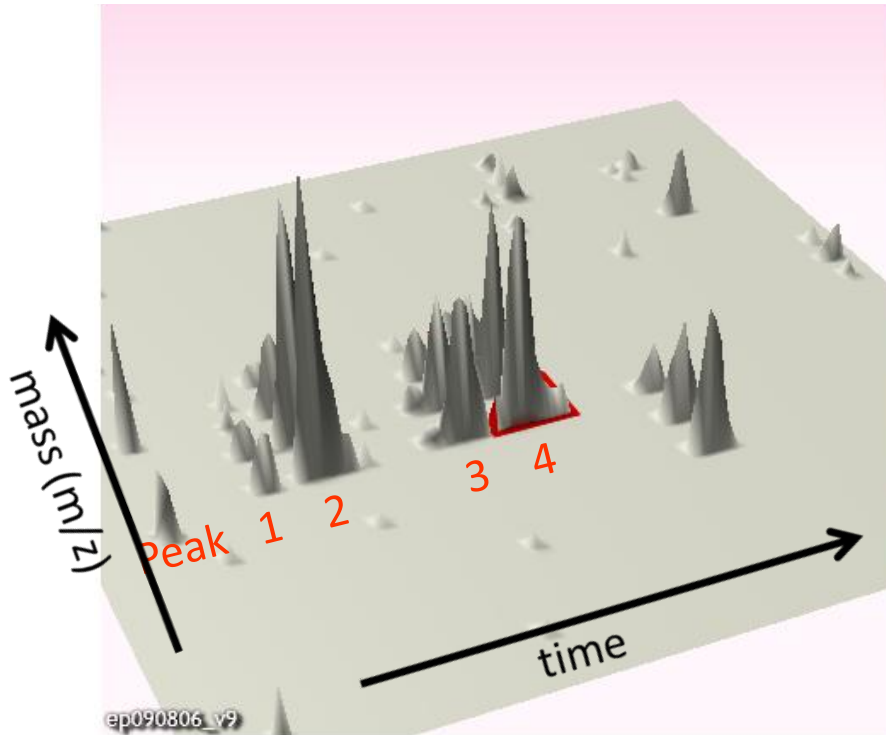


# Extracted ion chromatograms of SKGDSFSSSIPLSTK

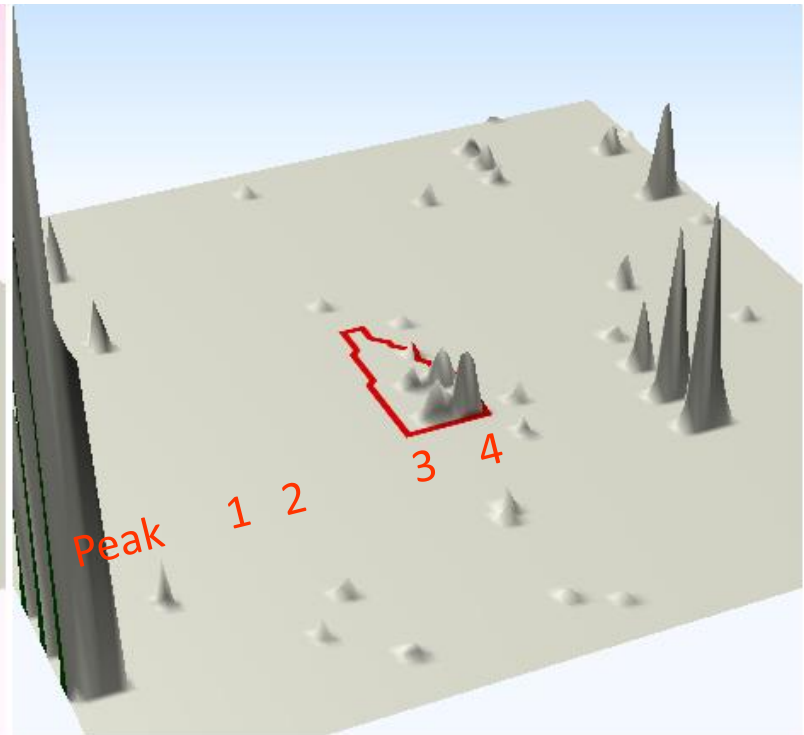


# Peak areas of SKGDSFSSSIPLSTK phosphorylated

elicited with chitin



Not elicited with chitin

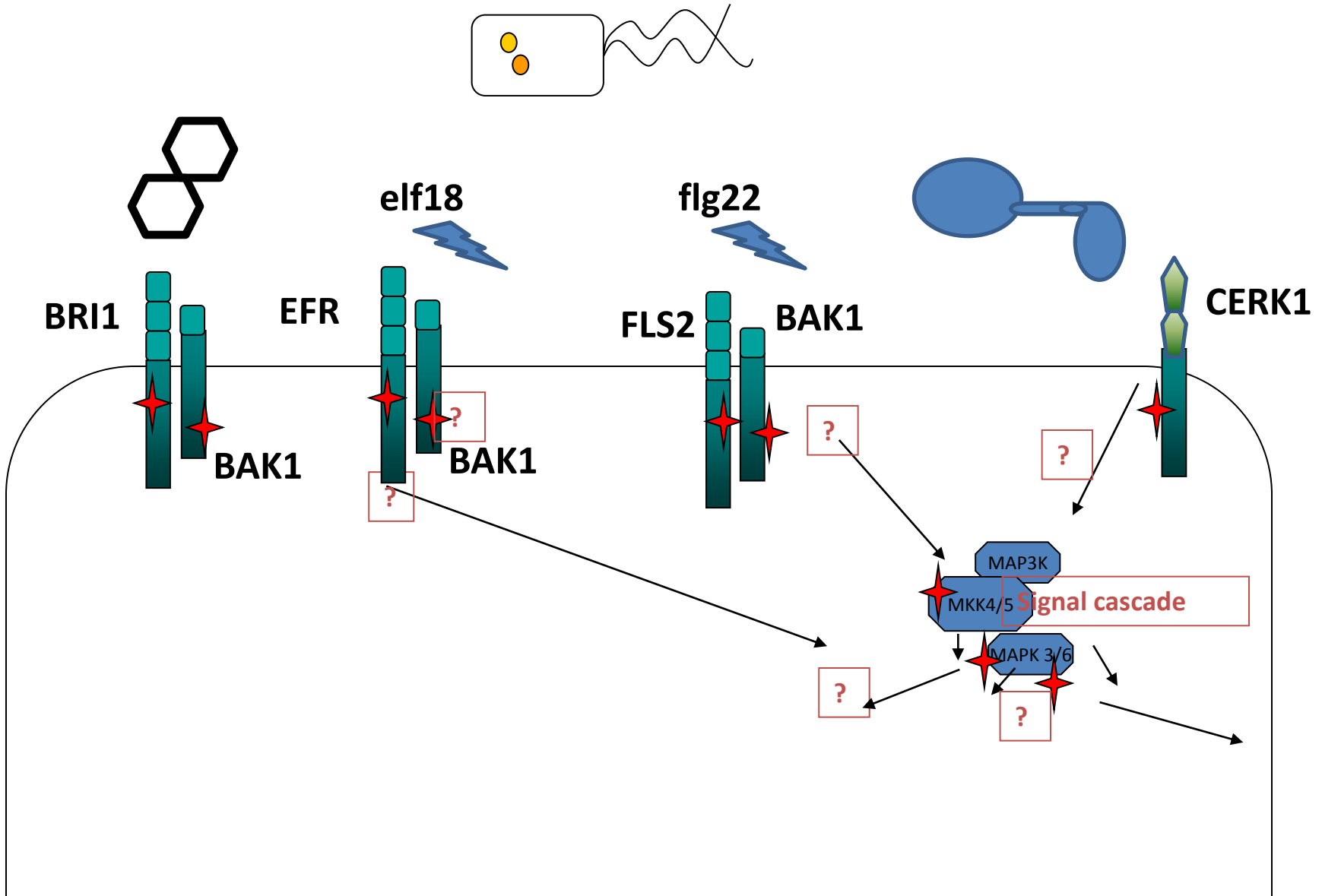


# Constitutive and induced phosphorylation revisited

Peptide	modification	percent of specific forms of peptide							
		plus chitin				no chitin			
		B1T1	B1T2	B2T1	B2T2	B1T1	B1T2	B2T1	B2T2
GDSFSSSIPLSTK		34	36	35	32	40	30	39	47
GDSFSSSIPLSTK	Phospho (S274)	0	0	1	1	0	0	-	0
SKGDSFSSSIPLSTK 2+	Phospho (S274)	1	2	4	4	0	0	0	0
SKGDSFSSSIPLSTK 2+	Phospho (S266)	2	1	4	3	2	1	2	2
SKGDSFSSSIPLSTK 2+	Phospho (S268)	3	4	3	3	1	1	0	0
SKGDSFSSSIPLSTK 2+		35	34	37	40	32	37	39	33
SKGDSFSSSIPLSTK 3+		25	24	17	17	25	30	19	17
GAVVKMTEAVGEFR	Oxidation (M)	81	71	44	36	100	99	100	100
GAVVKMTEAVGEFR	Oxidation (M)								
GAVVKMTEAVGEFR	Phospho (T519)	19	29	56	64	-	1	-	-

**B= biological replicate, T= technical replicate**

# The co-receptor kinase BAK1



# Current BAK1 strategy for mapping and quantifying phosphorylation sites

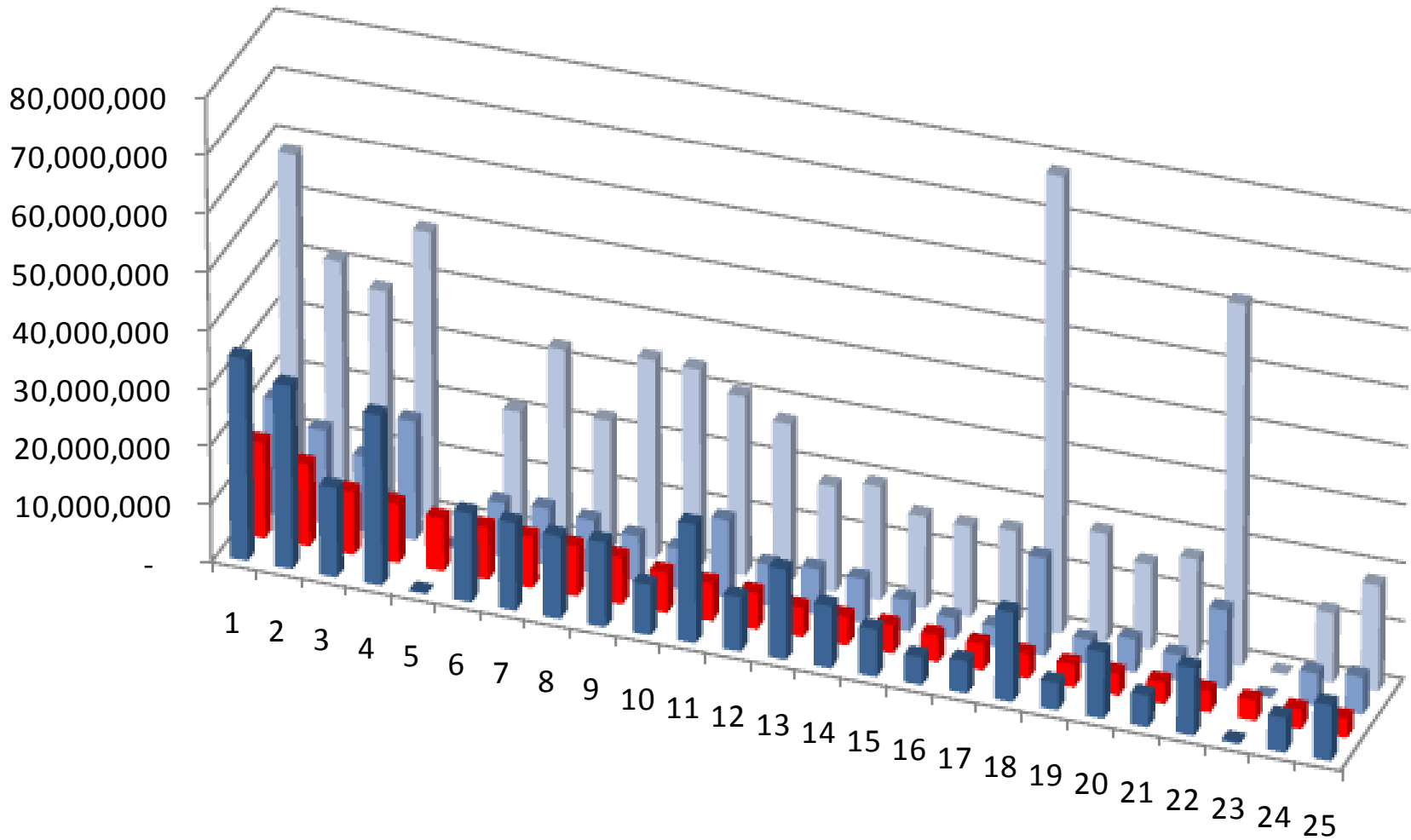
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Expressed, tagged proteins

WT, hyperactive mutant, kinase dead versions

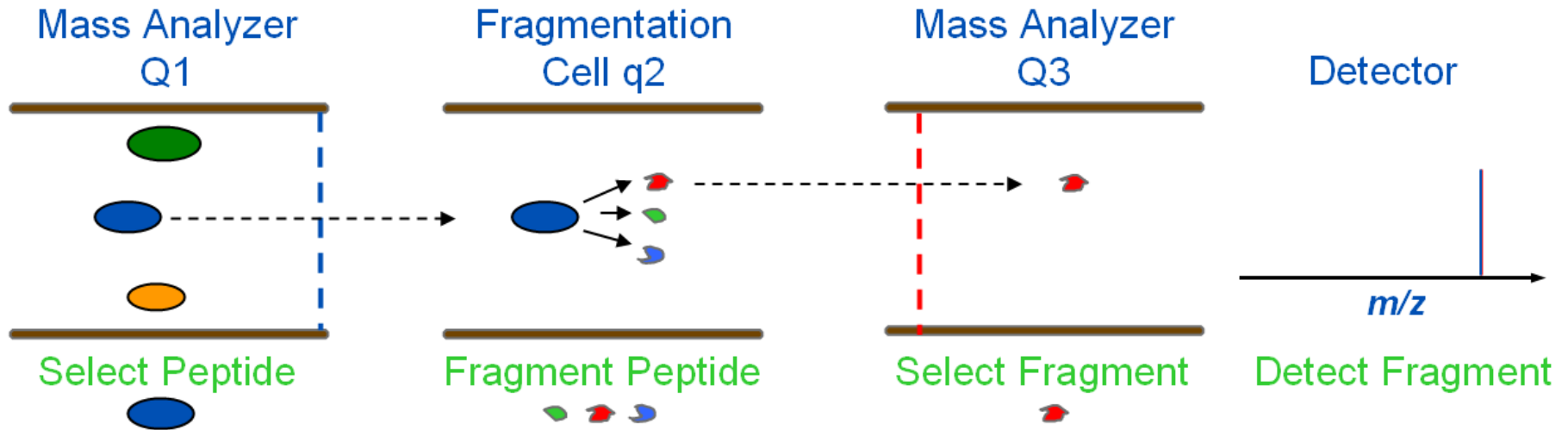
1. In vitro: 'Hot + cold' kinase assays  
Gel-LC-MS/MS mapping
2. In vivo: IP from transient expression *N. benthamiana*,  
IP stable transgenics *Arabidopsis*
3. In vivo: native promoters, SRM

# Peak areas of top 25 BAK1 peptides





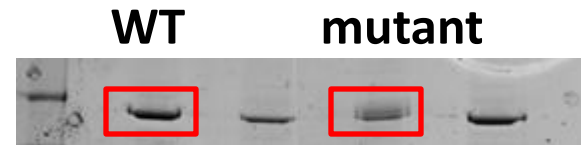
# Multiple reaction monitoring



## Note:

Modified forms of a peptide differ in both Q1 (intact mass) and Q3 (fragment mass)

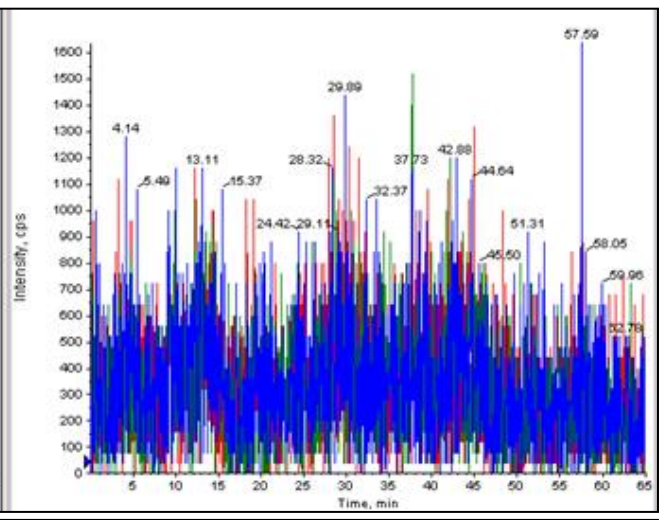
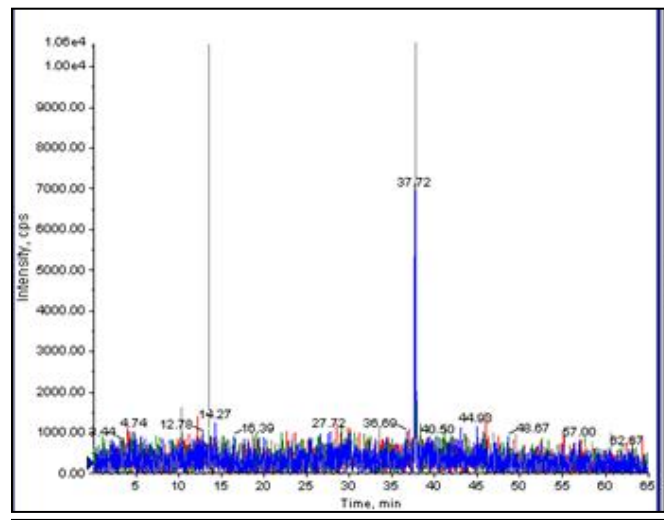
# Phosphorylation events on BAK1



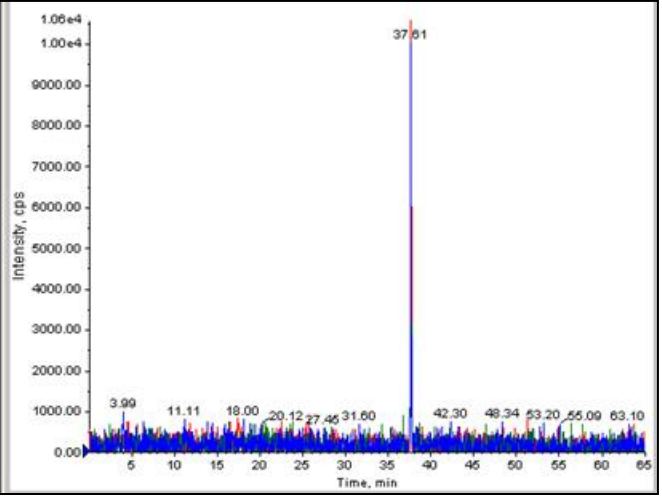
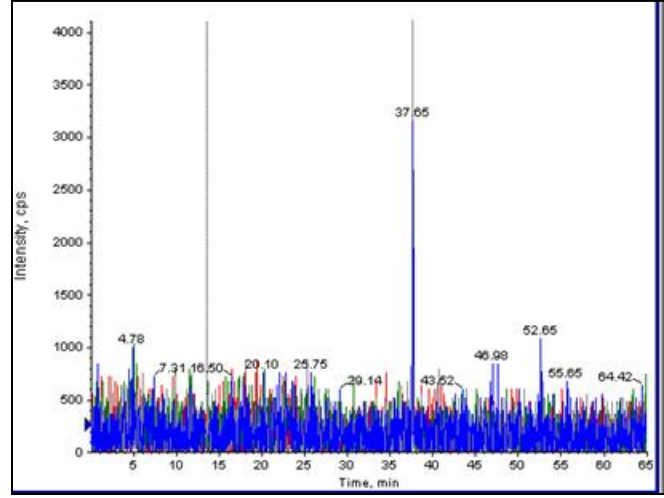
Non-phosphorylated

phosphorylated

WT



mutant

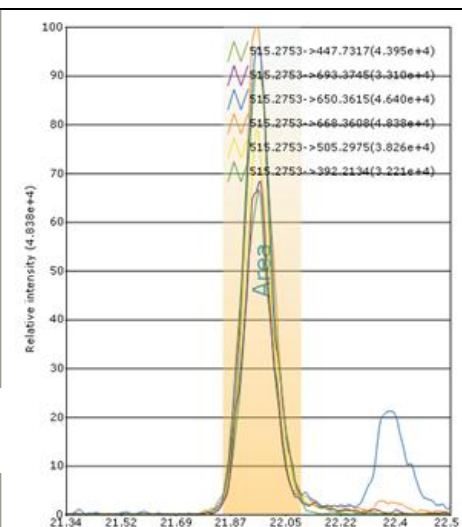
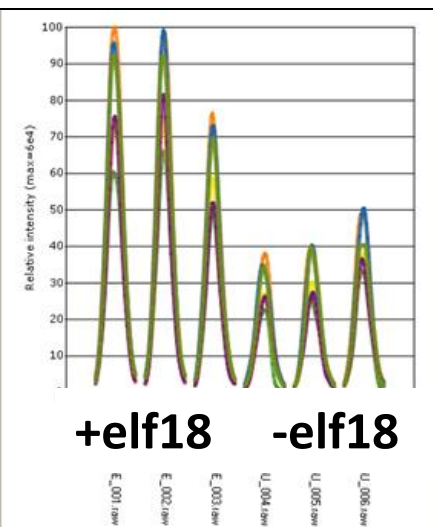
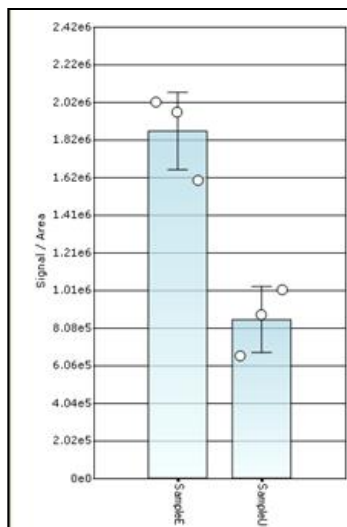


in-house Qtrap 4000

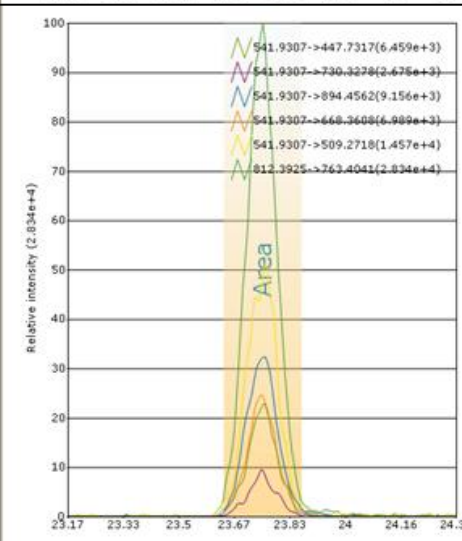
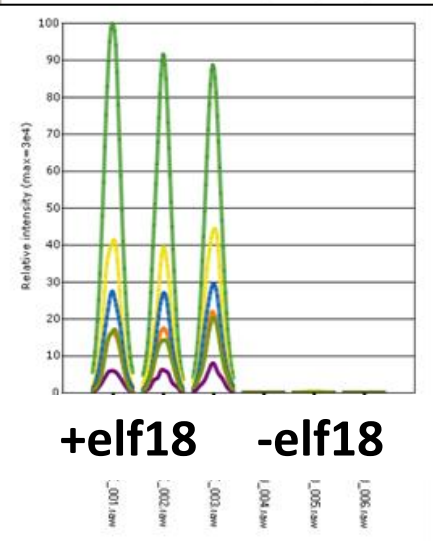
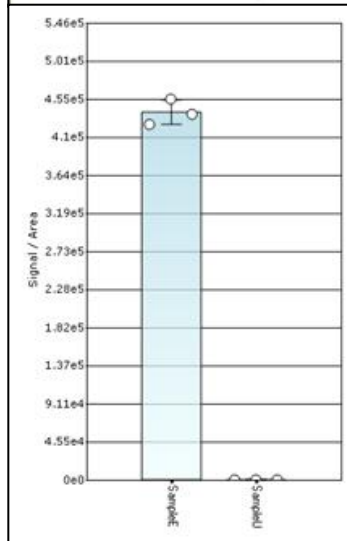
Analyst image, AB Sciex

# Phosphorylation events on BAK1 with elicitors

Non-phosphorylated



phosphorylated



## Summary

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- Plant proteomics is highly feasible
- Phosphorylation is important in plant-pathogen interactions
- Identification of sites is becoming routine

Current challenges are

assignment of correct sites\*  
quantification

- Future challenges  
subcellular localisation,  
composition of complexes

\*Note biochem noise, possible flexibility of enzymes

## **CERK1**

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Ulrike Lipka  
Volker Lipka

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Antje Hesse-Peck,  
Vardis Ntoukakis,  
Tatiana Mucyn,  
John Rathjen

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Cyril Zipfel

## **MS group**

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Jan Sklenar

